## Synthetic Studies of Antitumor Natural Products Superstolides A and B. Construction of C20–C26 Fragment of Superstolide A

2001 Vol. 3, No. 10 1447–1450

ORGANIC LETTERS

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Received February 14, 2001

## ABSTRACT





The C20–C26 portion of the antitumor macrolide superstolide A was synthesized by employing Brown's asymmetric crotylboronate methodology.

Superstolides A (1) and B (2) were isolated from the deep– water marine sponge *Neosiphonia superstes* collected off New Caledonia.<sup>1</sup> The structural novelty of these two molecules is characterized by a unique 16-membered macrolactone attached to a highly functionalized *cis*-decalin (Figure 1).

Superstolides A (1) and B (2) are highly cytotoxic against human bronchopulmunary non-small cell lung carcinoma NSCLC-N6-L16 cells with IC<sub>50</sub> values of 0.04 and 0.039  $\mu$ g/mL, respectively.<sup>1</sup> Both superstolides A (1) and B (2) exhibited potent cytotoxicity against murine leukemia P388 cells with an IC<sub>50</sub> of 0.003  $\mu$ g/mL and human nasopharyngeal carcinoma KB cells with IC<sub>50</sub> values of 0.02 mg/mL and 0.005  $\mu$ g/mL, respectively.<sup>1</sup> In addition, superstolide A (1) is also highly cytotoxic against human colon carcinoma HT29 cells with an IC<sub>50</sub> of 0.04  $\mu$ g/mL, and murine leukemia cells



## Figure 1.



expressing resistance toward doxorubicine P388 Dox with an IC<sub>50</sub> of 0.02  $\mu$ g/mL.<sup>1</sup> Roush and co-workers published a diastereoselective synthesis of the *cis*-fused decalin using an intramolecular Diels—Alder reaction in 1996.<sup>2</sup> However, a complete total synthesis has not yet been reported.

As part of our program studying the chemistry and biology of antitumor natural products, we recently initiated a project directed toward total synthesis of superstolides A and B. Our retrosynthetic analysis of superstolide A (1) is shown in Scheme 1. Disconnections at C2–C3, C6–C7, and C19– C20 reveals three key fragments, **3**, **4**, and **5**, with Horner– Emmons olefination, Julia olefination, and Suzuki (or Stille) coupling playing crucial roles in the synthetic strategy. Herein, we wish to report the asymmetric synthesis of fragment **4**, the linear polypropionate (C20–C26) portion of superstolide A.

(S)-(-)-Ethyl lactate **10** was the point of departure for the synthesis of the key fragment **4** (Scheme 2). Compound **10** 



reacted with *p*-methoxybenzyltrichloroacetimidate under acidic conditions to give compound **11** in 91% yield.<sup>3</sup> Reduction of ester **11** with DIBAL at -98 °C provided (*S*)-2-(4-methoxybenzyloxy)propionaldehyde **9** in 89% yield. Reaction of aldehyde **9** with (*d*)-(*E*)-crotyldiisopinocampheylborane **12** under Brown's conditions<sup>4</sup> afforded homoallylic alcohol **8** with complete diastereo- and enantioselectivity. Protection of the secondary alcohol **8** by TBSOTf afforded compound **13** in 64% yield from **9**. The oxidative cleavage of the double bond of compound **13** under the literature conditions<sup>5</sup> gave the desired aldehyde **7** in low yield (60–64%) due to the overoxidation of the diol intermediate leading to the formation of  $\alpha$ -hydroxy ketone **14** (Scheme 3). We found the reaction solution to be acidic (pH = 2).



Addition of a base could suppress the formation of compound **14** and increase the yield of aldehyde **7**. After screening of a few bases, we found that 2,6-lutidine gave the best results without racemization of the  $\alpha$ -methyl group. The desired aldehyde **7** was obtained in 90%.

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Aldehyde **7** reacted with (*l*)-(*E*)-crotyldiisopinocampheylborane **15** under Brown's conditions<sup>4</sup> to provide homoallylic alcohol **16** along with minor diastereomer **17** (6:1) (Scheme 4). The stereochemistry of the secondary alcohols of compounds **16** and **17** was determined on the basis of the <sup>13</sup>C NMR analysis of 1,3-diol acetonides **18** and **19** derived from **16** and **17**, respectively.<sup>6</sup> It is surprising that the configuration of the secondary alcohols in both **16** and **17** is the requisite *R*. The absolute configuration of the methyl groups at C22 of compounds **16** and **17** was determined to be *R* and *S*, respectively, on the basis of proton homonuclear decoupling analysis, NOESY experiments, and <sup>13</sup>C NMR analysis of 1,3-diol acetonides of **20** and **21**.

After alcohol **16** was protected by a TES group, the *p*-methoxybenzyl group was removed by DDQ to give compound **6** in 94% yield (Scheme 5). The attempt to convert **6** to **23** under Mitsunobu conditions with diphenyl phosphorazidate<sup>7</sup> led to the exclusive formation of an intramolecular silyl transfer product. Our initial effort to employ  $HN_3^8$  as the azide source gave either mainly intramolecular silyl transfer product or simply no reaction.<sup>9</sup> However, we found that by employing freshly prepared  $HN_3$  which was further dried over 3 Å MS, the Mitsunobu reaction went smoothly and the desired azide **23** was obtained in 84% yield. Reduction of the azide **23** by Staudinger reaction<sup>10</sup> followed by acylation of the primary amine afforded amide **24** in 84% yield from **23**. Oxidative cleavage of the terminal double bond of **24** under our modified procedure gave aldehyde **25** in 71% yield. By employing Takai's reaction, aldehyde **25** was homologated to the requisite *trans*-vinyl iodide **4** in 71% yield with a ratio of 11:1 of *trans* to *cis* isomer.<sup>11</sup>



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In conclusion, fragment 4 was successfully synthesized in 12 steps using Brown's asymmetric crotylborane methodologies. By employing the enantiomer of starting material 10 and different crotylboranes, it will be possible to synthesize different diastereomers and enantiomers of fragment 4 which will be used in SAR studies. Currently, the asymmetric synthesis of fragment 3 is underway and will be reported in due course. Acknowledgment. This work was financially supported by a Research Project Grant RPG-00-030-01-CDD from the American Cancer Society, Grant IN-122S from the American Cancer Society, administered through The University of Iowa Cancer Center, the Central Investment Fund for Research Enhancement (CIFRE) of the University of Iowa, and a fellowship from The Center for Biocatalysis and Bioprocessing at The University of Iowa (to W.Y.).

**Supporting Information Available:** Complete description of experimental details and product characterization. This material is available free of charge via the Internet at http://pubs.acs.org.

OL0100273

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